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Gastrointestinal nematodes infections in ruminants: Prevalence and

Anthelmintic efficacy

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Abstract:

This study investigates the prevalence of gastrointestinal nematodes in ruminants in Ismailia, Port-said and Damietta cities, during a period from October 2016 to September 2017. A total of 130 fecal samples were collected from sporadic cases and 60 abomasal samples and 36 small intestinal samples were collected from slaughtered animals in abattoirs to detect the adult nematode parasites. The results indicated that sheep was infected with 5 species of nematodes 3 in abomasum and 2 types in small intestine. The prevalence of the infection were 26.15%, in faecal, abomasal. and small intestine samples 30% and 11.11% respectively. The highest prevalence of the nematodes was in young animals (71.42% in ovine sp., 14.28% in bovine sp. and 0.0% in caprine sp). Higher prevalence was in male sheep 60% than ewe 30%. Damietta was the highest in prevalence (47.6%) then in Ismailia (29.16%) and 0.0% in Port-said. The most prevalent species of nematodes in abomasum was Haemonchus contortus (26.67%), Ostertagia sp. (10%) and Trichostrongylus axei (3.33%). In small intestine, were Cooperia sp. (11.11%) and Nematodirus sp. (5.56%). Faecal egg count reduction test was carried out on Ivomec super & Albendazole to demonstrate the efficacy of anthelmintics, and proved the Ivomec was more potent than Albendazole in treatment of GINs.

Key words Ruminants-Nematodes-H. contortus- Histopathological changes

Introduction:

Ruminants are considered a major source of meat and milk productions for human consumption Egypt. in Egypt possesses large population of livestock which is managed by smallholder farmers.Gastrointestinal nematode infection was one of the most problems on health all over the world, it was affected the health of several people and

animals causing huge economic loss in ruminants farming (Muluneh *et al.*, 2014).

Haemonchus was regarded as one of the most prevalent and highly pathogenic, possessed the highest biotic potential and had a prominent ability to develop resistance against most widely used anthelmintics and a unique survival strategy due to great biological and ecological plasticity. Hence compared to gastrointestinal other nematodes. Haemonchus was the most important parasite of domestic ruminants especially in sheep and goats (Kumsa et al., 2008).

Anthelmintic treatment considered the most common method to control of GINs in ruminants although the cost of medication and anthelmintic resistance was a good documented phenomenon (EI-Gayar, 2002). Currently, the primary control strategy for nematode infections was the use of chemical treatments (Ahmed, 2010).

Material and methods:

1. The Study area & animals:

This study was carried out on ruminants animals (cattle, buffalo, sheep and goats) which are collected from Ismailia, Port-said and Damietta. 130 faecal samples (64 from sheep, 18 from goat, 34 from cattle, 14 from buffalo) were collected from lived animals, sixty abomasal (54 from sheep, 4 from cattle and 2 from buffalo) & 36 small intestinal (32 from sheep, 2 from cattle and 2 from goat) samples were collected from abattoirs. The sex and age of studied animals were recorded. Parasitological examination were done in Faculty of Veterinary Medicine, Suez Canal University.

2. Laboratory procedures: 2.1. Faecal examination:

(A) Macroscopic examination: Each faecal sample was inspected by the naked eyes and with the aid of a magnifying hand lens for the presence of any gross parasites. Blood or mucous, colour, odour and consistency of the faeces were recorded.

(B) Microscopic examination:
Concentration flotation technique:
i. Saturated Salt Centrifugal fecal floatation technique according to (Zajac and Conboy, 2012):

2.2 Abattoir samples examinations:

(A) Worm counts: Following slaughter, the abomasum and small intestine of each animal removed, the abomasum was opened along its greater curvature and the content collected in a container. Recovery and counting of worms from the abomasum and small intestine were carried out using similar technique to that described by Bissat et al., 1996. nematode worms The were collected, preserved, identified and counted (Demissie et al. 2013 and Vadleich et al. 2014). Complete identification of the nematodes was carried out after permanent mount acc. to Glycerol Jelly Method: (Fleck & Moody, 1993)

2.3 Histopathological examination:

(A) Tissue samples: For histopathologicl examination, abomasal tissues collected from slaughtered animals were sectioned. Sections of adjacent grossly uninvolved tissue were also collected. The sections were fixed in 10% neutral buffered formalin.

(B) Histopathological studies: Abomasum specimens were fixed in 10% neutral buffered formalin for 72 hrs. Tissue samples of 1 to 2 mm thickness were dehydrated in graded alcohol and cleared in xylene and embedded in paraffin blocks. The 4-5µ thick serial sections were taken with rotator microtome on clean grease free slides processed and stained with Haematoxyline & Eosin stain (Luna, 1968).

3. Evaluation of the efficacy of some anthelmintics by using faecal egg count reduction test (FECRT):

- (A) Animals used for the evaluation of the efficacy of the selected anthelmintics:
- Ten of identified infected animals were detected by concentration flotation technique as they were positive for *Strongyle*-type eggs were selected for evaluation of the efficacy of Ivomec super (Ivermectin, Clorsulan) and Albendazole (Albendazole 2.5%).
- All of the studied infected ruminants were divided into two similar groups, each of 5 animals which used for treatment trial.
- All animals (10) in both group were examined pretreatment for *Strongyle*-type eggs which were counted using McMaster egg-count technique (pretreatment count).
- The first group (5animals) was treated by Ivomec super (Ivermectin, Clorsulan) while the second group was treated by Albendole (Albendazole 2.5%).

(B) The selected anthelmintics:

Table (1): Anthelmintics generic name, route of administration and recommended dose

Trad e name	Generic name	Route of administr ation	Recommen ded dose
Ivom ec Supe r	Ivermecti n, Clorsulan	subcutane ous injection	1 ml / 50 kg bodyweight
Alben d- azole	Albendaz ole 2.5%	oral drench	10ml /50 kg. bodyweight

(C) Egg counting technique:

Egg count was determined for positive samples only using the McMaster technique according to **Soulsby (1982)**.

No. of eggs/gram = $\frac{\text{Total No.of eggs counted}}{\text{No.of counting chamber}} \times 200$

(E) Efficacy %: (Knox, 2002): Efficacy Reduction of FEC % =

 $\frac{\text{pre-post}}{\text{pre}} \times 100$

Results:

In the current study on the aastrointestinal parasites in ruminants in Ismailia, Port-Said and Damietta governorates, out of 130 faecal samples of ruminants animals examined (64 from sheep, 18 from goat, 34 from cattle, 14 from buffalo) 34 samples (26.15%) were found to be positive for the aastrointestinal nematodes eaa and out of 60 abomasal samples (54 from sheep, 4 from cattle and 2 sample from buffalo) 18 samples (30%) were found to be positive for the abomasal samples andout of 36 small intestine samples (32 from sheep, 2 sample from cattle and 2 sample from goat) 4 samples (11.11%) were found to be positive for the small intestine samples of different age, sex, species and origin of animals were examined. The detected nematodes were (Haemonchs, Ostertagia, Trichostrongylus, Cooperia and Nimatodirus) were recovered.

1. The prevalence:

Table (2): Prevalence of (GINs) in ruminants in relation to age of the animals

	Age	No. of	Prevalence (%)			P-value				
Samp le Spp.		Faecal sample	Abomasal sample	S.I. sample	Fae.	Ab	S.I.	Fae.	Ab.	S.I.
Bovine	Young Adult	4/28 0/20	0/4 0/2	0/2 0.0	14,28 0.0	0.0 0.0	0.0 0.0			
Ovine	Young Adult	30/42 0/22	18/36 0/18	4/24 0/8	71,42 0.0	5 0.0	16.67 0.0	0,008	008. •	0,54
Caprine	Young Adult	0/10 0/8		0.0 0/2	0.0 0.0		0.0 0.0			

Table (3): Prevalence of GINs in ruminants in relation to sex of the animals

	Sex	No. of infested/ No of examined			Prevalence (%)			P-value		
Samples Sp.		Faeca I sampl e	Aboma sal sample	S.I. sample	Fae.	Ab.	S.I.	Fae.	Ab.	S.I.
Bovine	Male Female	2/22 2/30	0/6 0.0	0/2 0.0	9.1 6.67	0.0	0.0	0.83		
Ovine	Male Female	24/40 6/20	14/46 4/8	4/30 0/2	60 30	30.43 50	13.33 0.0	0.055	0.498	0.058
Caprine	Male Female	0/6 0/12		0.0 0/2	0.0 0.0	-	0.0 0.0			

Fig. (1): Monthly prevalence of GINs

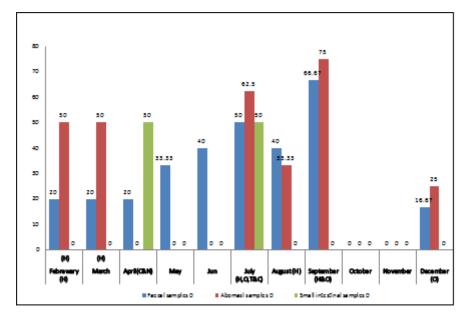


Table (4): The prevalence of GINs in relation to location of the animals

2. Morphological features:

	No. of infested/ No of examined Prevalence (%)						P-value		
Sample Location	Faecal sample	Abomas al sample	S.I. sample	Fae.	Ab.	S.I.	Fa e.	Ab.	S.I.
Ismailia	14/48	4/20	2/12	29.16	20	16.67	0.0	0.000	0.44
Port-said	0/40	0/12	0/10	0.0	0.0	0.0	0.0 00	0.003 3	0.41 3
Damietta	20/42	14/28	2/14	47.6	50	14.28	4		



Fig. (2): Strongyle-type egg

Fig. (3): Haemonchus contortus anterior en X10



Fig. (4): Haemonchus contortus male spicules and Y shaped dorsal ray X10



Fig. (5): Haemonchus contortus female white uterus winding around the red blood-filled intestine giving a twisted or barber`s pole appearance X4

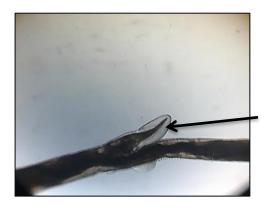


Fig. (6): *H. contortus* female vulvar flap, linguiform type (black arrow) X4

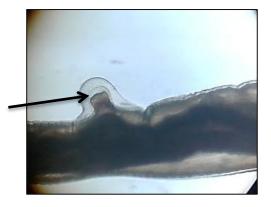


Fig. (7): Haemonchus contortus female vulvar flap, knob type (black arrow) X4



Fig. (8): Nematodirus sp. anterior end with cephalic swelling (black arrow) X10



Fig. (9): *Nematodirus* sp. male bursa (a) and spicules (b) X4



Fig. (10): *Nematodirus* sp. female vulvar region (black arrow) X20

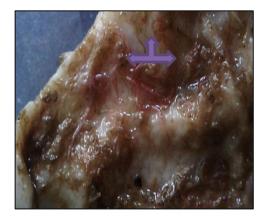


Fig. (11): Portion of abomasum with adult *Haemonchus* worms (arrows)

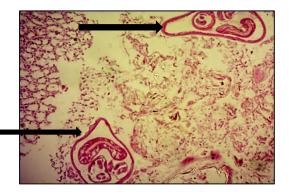


Fig. (12): Sheep, abomasum showing two cross sections of *Haemonchus contorus* (black arrow) with massive destruction and desquamation of the superficial layer of mucosa in the lumen. H&E, X 200.

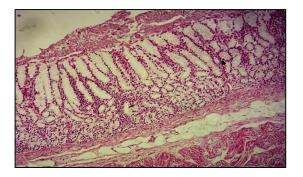


Fig. (13): Sheep, abomasum showing necrosis of the superficial layer of mucosa, diffuse mucinous degeneration of submucosal glands with leukocytic infiltrations. H&E, X 200.

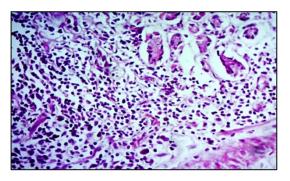


Fig. (14): Sheep, abomasum higher magnification of previous figure showing massive necrosis of the glands which was replaced by infiltrations leucocytic mainly lymphocytes, macrophages and few plasma cells. H&E, X 400.

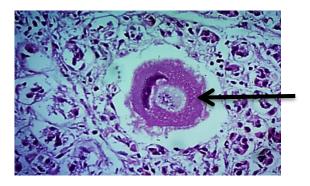


Fig. (40): Sheep, abomasum higher magnification of previous figure showing larva of *ostertagia* in gastric gland (black arrow) gland (black arrow) X 400.

4. Evaluation the efficacy of some anthelmintics by using faecal egg count reduction test (FECRT):

Efficacy of used anthelmintics on GINs infesting the selected group of sheep:

Table (5): Changes in mean egg per gram in Ivomec super (Ivermectin, Clorsulan) treated sheep group.

		Changes in	Changes in mean egg/gram in examined sheep									
	Animal	Animal Before 1st wk 2nd wk after		3rd wk	4th wk							
	number	treatment	after ttt	ttt	after ttt	after ttt						
Treated	1	1500	200	100	100	100						
group	2	1000	200	100	0	0						
	3	500	0	0	0	0						
	4	700	100	0	0	0						
	5	1200	200	100	0	100						
Mean		980	140	60	20	40						
egg /gm												
Efficacy			85.71	93.87	97.95	95.9						
%												

Table (6): Changes in mean egg per gram in Albendazole (Albendazole 2.5%)

treated sheep group.

		Changes in mean egg/gram in examined sheep								
	Animal	Before	1st wk	2nd wk	3rd wk	4th wk after				
	number	treatment	after ttt	after ttt	after ttt	ttt				
Treated	1	500	0	0	0	0				
group	2	400	0	0	0	0				
	3	1700	200	200	100	200				
	4	900	100	0	0	100				
	5	1300	200	100	100	200				
Mean		960	100	60	40	100				
egg /gm										
Efficacy			89.58	93.75	95.83	89.58				
%										

Discussion:

1. Prevalence status:

The prevalence of gastrointestinal nematodes infection were 26.15%, 30% and in faecal, abomasal. and 11.11% small intestine samples respectively The current results were higher than those reported by Eisa & Esmail (2003) (28.9%), Abouzeid et al. (2010) (27.5%), Khalafalla et al. (2010) (10.4%), Awraris et al. (2012) (27.57%), Kabaka et al. (2013) (13.8%), Mir et al. (2013) (21.93%), Swapna and Nithinya (2015) (19.36%) and Sultan, et al., (2016) (26.79%). The fluctuation in prevalence of GINs might be due to extensive use of anthelmintics by the farmers, difference in agro-climatic conditions that could support prolonged survival and development of infective larval stages most nematodes of (Muluneh et al., 2014).

In the present study, the age susceptibility of the examined animal infected with GINs was observed all over the year with the highest percentage of infection and significantly high in young animal (2-6 months) but the infection was nil in adult animals (more than one year old). The high prevalence of GINs in young animal than adults may be due to the lack of immunity which acquired by continuous infection due to bad management especially space insufficiency and continuous indiscriminate treatments which only minimizes the intensity of infection without efficient integrated control measures (Lemma and Abera, 2013). This result was similar with that recorded by Regassa *et al.* (2006), Pfukenyi *et al.* (2007), Abouzeid *et al.* (2010), Awraris *et al.* (2012), Kakar *et al.* (2013), Lemma and Abera (2013), Lashari *et al.* (2015), Swapna and Nithinya (2015).

The present study revealed that of the animals sex shows significance variation at (p≤0.05) with GINs infection in case of faecal samples of sheep as the prevalence of infection in males was 60% and in females was 30%, this may be due to stimulatory effects of estrogens and inhibitory effect of androgens on immune responses (Lashari et al., 2015) and (Desta, 2015).

The study further revealed that the highest prevalence of GINs egg was during September (66.67%) followed by July (50%) then in months (June and August) (40%) then in May (33.33%) and then in months (February, March and April) (20%), the lowest were found in (16.67%) and December with (0.0%) in the other months. Also adult worms in abomasal samples were higher in September (75%) then in July (62.5%) then in February and March (50%) then in August month (33.33%), and the lowest prevalence was recorded in December (25%) and (0.0%) in other months. These results were agreed with Nwosu et al. (2007) and Mir et al. (2013) as their results recorded that Haemonchus and Trichostrongylus species

attained peak counts together in both goats (June) and sheep (August).

Concerning adult worms in abomasum, 30% of the examined abomasal samples were infested with at least one species of nematodes worms. The most prevalent species was Haemonchus contortus (26.67%) then Ostertagia sp. (10%) and Trichostrongylus axei (3.33%) from the examined abomasal samples, and Cooperia sp. (11.11%) and Nematodirus sp. (5.56%) from examined small intestine samples. These results were in line with results of Raza et al. (2009) as the prevalence of *H. contortus* in sheep 37.18% in all examined was animals. Sultan et al. (2010) in Gharbia Governorate, Egypt, Owhoeli al. (2014) et and Varadharajan and Vijayalakshmi (2015) recorded that the most prevalent abomasal nematode was H. contortus.

The present study revealed that location of the animals showed high significance variation so in of faecal and case abattoir samples. The highest prevalence of nematodes were recorded in Damietta (47.6% - 50%) then Ismailia (29.16% - 20%) and with (0.0%) in Port-said city. In our opinion the absence of infection in Port-said city may be due to the good managemental activities of the farmer such as housing of the animals. good feeding and periodical using of anthilmintics, similar findings were observed by Kabaka et al. (2013).

2.Morphological status:

The morphological status including that of Haemonchus contortus recovered from 16 abomasal samples of the sheep. The male worms were reddish in color while the females were with white uterus which spirally wound around the red intestine, giving the appearance of a barber's pole. The male length was 12.6-19.4mm, the dorsal lobe of the bursa was supported by Y shaped dorsal ray and the spicules were barbed. The female was 14.4-25 mm in length, in most of the specimens, the vulva was covered with a conspicuous large flap, the flap was linguiform, in a few specimens it was reduced to small knob, this feature were agreed with finding of Lichtenfels et al. (1986 & 1994), El-Gayar (2002), Rauf Tak et al. (2014) and Saminathan et al. (2015).

In case of description of Ostertagia sp. which was collected from the 6 abomasal samples of sheep. The male measured 6.2-7.5 mm long. the spicules were pigmented brown, equal, short and trifurcated distally. The female length of Ostertagia was 8.1-9.2 mm and the female tail was slender and curved in a C-shape manner. These morphological characters were similar to those described by Lichtenfels and Pilitt (1991) and El-Gayar (2002).

Trichostrongylus axei were collected from 2 abomasal samples of sheep. The worms were small, Cervical papillae were lacking. The male was 2.5-6 mm long and the female was 3.5-8mm long. The spicules of male were dark brown in color, and unequal. This description was similar with those obtained by **Boomker (1986)**, **Knight (1962), Soulsby (1982)** and **Ghasemikhak** *et al.* (2011).

In the present study, *Nematodirus* sp. were collected from 2 specimens small intestine of sheep. They have an inflated cuticle around the anterior end with 14-18 longitudinal ridges on the cuticle. This result was similar to that obtained by **Lichtenfels and Pilitt (1983)**, **Hoberg et al. (1986)** and **Hoberg and Rickard (1988)**.

Cooperia sp. were collected from 4 specimens of small intestine. The cuticle of the anterior end formed a cephalic swelling with transverse striations and the rest of the body cuticle beard 14-16 longitudinal ridges. The female was 5.7-7.5 mm long. This description was similar to that recorded by **Lichtenfel (1977)** and **Soulsby (1982).**

3.Histopathological findings:

In gross lesions, there were a watery content of the abomasum which partially covered with free blood. Numerous adult worms and petechial hemorrhage were found on the abomasal mucosa. This result was agreed with **Tehrani** *et al.* (2012) and **Saminathan** *et al.* (2015).

Current study referred to histopathological changes on the examined abomasum of sheep which showed cross section of *Haemonchus contorus*. The worms was seen embedded between mucosa and submucosa with destruction massive and desguamation of the superficial layer of mucosa that lead to focal necrosis of the superficial layer of mucosa. Submucosal layer was leukocytic showed prominent infiltrations. This result was agreed with that obtained by El-Gavar (2002) and AL-Hasnawy (2014).

4. Efficacy of anthelmintics:

the present study, the In anthelmintics efficacy of two Ivomec super (Ivermectin) and Albendazole in treatment of gastrointestinal nematodes naturally infecting animals was evaluated by using faecal egg count reduction test (FECRT). Results revealed that Ivomec super was more efficient as it induced marked reduction in mean egg /gm from which was 980 before treatment to become 40 in the 4th week after treatment. The means epg of this group were (980) pretreatment and (140, 60, 20 and 40) with post-treatment efficacy 85.71%, 93.8%, 97.95% and 95.9% in 1st, 2nd, 3rd and 4th week after treatment respectively. This finding was nearly similar to that obtained Mckellar et al. bv (1991). Moreover Goudie et. al. (1993), Walsh et. al. (1995) and Khaled et al. (2004).

Conclusion:

From this study, it could be concluded that:

- Ovine gastrointestinal nematodes were the major helminthosis parasites in ruminants.
- The age, monthly variations and the location of the animals were very important factors associated with gastrointestinal nematode infection.
- The most prevalent type of gastrointestinal nematode was *Haemonchus* sp. which cause severe pathogenic effect in infected animals.
- ✤ Ivomec super as an anthelmintic was more effective than Albendazole in treatment of GINs. SO it was recommended to treat infected ruminants with this anthelmintic in months with high prevalence (June, July, August and September).

References:

Abouzeid, N.Z.; Selim, A. M. and El-Hady, K. M. (2010): Prevalence of gastrointestinal parasites infections in sheep in the Zoo garden and Sinai district and study the efficacy of anthelmintic drugs in the treatment of these parasites. Journal of American Science, 6(11): 544-551.

M.A.A. Ahmed. (2010): Gastrointestinal (nematode) infections in small ruminants Epidemiology, anthelmintic efficacy and the effect of wattle tannins. Ph.M. School Agricultural of Sciences and Agribusiness, University of KwaZulu-Natal Pietermaritzburg.

Al-Hasnawy, M.H.M. (2014): Prevalence and pathogenicity of Haemonchosis in sheep and goats in Hilla city/Iraq. Journal of Babylon University/Pure and Applied Sciences, 22(7): 1989-1999.

Awraris, T.; Bogale, B. and Chanie, M. (2012): Occurance of gastrointestinal nematodes of cattle in and around Gondar Towen, Amhara regional State, Ethiopia. Acta Parasitologica Globalis 3(2): 28-33.

Bissat, S.A.; Vlassof, A.; Doch, P.G.C.; Jonas, W.E.; West, C.J. and Green R.S. (1996): Nematode burdens and immunological responses following natural challenge Romney lambs in selectively bred for low or high fecal worm egg count. Veterinary Parasitology, 61: 249-263.

Boomker, J. (1986): *Trichostrongylus auriculatus* N. SP. (Nematoda : *Trichostrongylidea*) from the Steenbok, Rapwcerus campestris (Thunberg, 1811). Onderstepoort Journal of Veterinary Research, 53: 213-215.

Demissie, T.; Tesfaye, D.; Fekadu, A. and Asefa, I. (2013): Study on abomasal nematodes of sheep and goats, comparison and characterization of vulvar morphology of *Haemonchus* in Hawassa, Ethiopia. African Journal of Agricultural Research, 8(39): 4922-4927.

Desta, H.M.G. (2015): Study on prevalence of GI nematodes in indigenous Bonga sheep breed at three selected Agro Ecologies of Kaffa and Bench Maji zones, Ethiopia. Journal of Biology, Agriculture and Healthcare, 5(13): 2224-3208.

Eisa, M.I. and Esmail, M. (2003): Concurrent gastrointestinal nematodes and psoroptic mange infestation in sheep: clinical investigation, some biochemical alteration and treatment. Veterinary Medical Journal, Giza, 51(2): 177-187.

El-Gayar, A.K. (2002): Morphological and immunological studies on gastrointestinal nematodes of goats. Ph.D. Thesis Faculty of Veterinary Medecin, Suze Canal University.

Fleck, S.L. and Moody, A. H. (1993): Diagnostic Techniques in Medical Parasitology. ELBS. with Buttetworth-Heinemann,

Publishers, London.

Ghasemikhah, R.; Mirhendi, H.; Gh.; Kia, E.B.: Mowlavi, Sarmadian. Η. ;Mehgi, B.: Golestan, B. and Mobedi, I. (2011): Morphological and morphometrical description of Trichostrongylus species isolated from domestic ruminants in Khuzestan province, Southwest Iran. Iranian Journal of Parasitology, 6(3): 82-88.

Goudie, A.C.; N.A.; Evans, A.F.: Bishop. Gration. B.F.: Gibson, S.P.; Holdom, K.S.; Kaye, Wicks, B.: S.R.; Lewis, D.: A.J.; Bruce, C.I,; Weatherley, Herbert, A. and Seymour, D.J. (1993): Doramectin — a potent novel endectocide. Veterinary Parasitology, 49(1): 5-15.

Hoberg, E.P. and Rickard, A.G. (1988): Morphology of the synlophe Nematodirus maculosus of (Trichostrongyloidea) with evolution comments on of Nematodirus spp. among the Caprinae (Artiodactyla). Helminthological Proceedings Society of Washington, 55(2): 160-164.

Hoberg, E.P.; Zimmerman, G.L. and Lichtenfels, J.R. (1986): First Nematodirus report of battus (Nematoda : *Trichostrongyloidea*) in North America: redescription and comparison to other species. Proceedings Helminthological Society of Washington, 53(1): 80-88.

Kabaka, W. M.; Gitaub, G. K.; Kitala, P. M.; Maingi, N. and VanLeeuwen, J. A. (2013): The prevalence of gastrointestinal nematode infection and their impact on cattle in Nakuru and Mukurweini districts of Kenya. Ethiopia Veterinary Journal, 17(1): 95-104.

Kakar, H.; Lateef, M.; Maqbool, A.; Abdul Jabbar, M.; Abbas, F.; Jan, S.; Abdul Razzaq; Kakar, E. and Shah, H. (2013): Prevalence and intensity of ovine gastrointestinal nematodes in Balochistan, Pakistan. Pakistan Journal of Zoology, 45(6): 1669-1677.

Khalafalla, R. E.; Elseify, M. A. and Elbahy, N. M. (2010): Seasonal prevalence of gastrointestinal nematode parasites of sheep in Northern region of Nile Delta, Egypt. Parasitology Research, 108(2): 337-40. Khaled, S.M.A.; Amin, M.R.; Mostofa, M.; Hossain, M.J. and Azad, M.A.K.A (2004): Effects of Vermic against gastro-intestinal nematodiasis in sheep. Journal of Biological Sciences, 4(6): 720-724.

Knight, R.A. (1962): Occurrence of Trichostrongylus Nematodes, longispicularis, Ostertagia lyrata, and Cooperia spatulata in ruminants in Mississippi. Mississippi Experiment Station Journal, 29(2): 145-147.

Knox, M.R. (2002): Effectiveness of copper oxide wire particles for *Haemonchus contortus* control in sheep. Australian Veterinary Journal, 80: 224-227.

Kudrnacova, M. and Langrova, I. (2012): Occurrence and seasonality of domestic sheep gastro-intestinal parasites. Scientia Agriculturae Bohemica, 43(3): 104-108.

Kumsa, B.; Tolera, A. and Abebe, R. (2008): Vulvar morphology and sympatry of *Haemonchus* species in naturally infected sheep and goats of Ogaden region, eastern Ethiopia. Veterinarski ARHIV 78(4): 331-342.

Lashari, M.H., Taswar, Z., Akhtar, M.S., Chaudhary, M.S. and Sial, N. (2015): Prevalence of Haemonchus contortus local in goats of D.G.Khan. World Journal of Pharmacy and Pharmaceutical Sciences, 4(05): 190-196.

Lemma, D. and Abera, B. (2013): Prevalence of ovine gastrointestinal nematodes in and around Asella, South Eastern Ethiopia. Journal of Veterinary Medicine and Animal Health, 5(8): 222-228.

Lichtenfels, J.R. (1977): Differences in cuticular ridges among Cooperia spp. of North American ruminants with an illustrated key species. to Proceedings of the Helminthological Society of Washington, 44(2): 111-119.

Lichtenfels, J.R. and Pilitt, P.A. (1983): Cuticular ridge patterns of Nematodirus (Nematoda : Trichostrongyloidea) parasitic in ruminants domestic of North America, with a key to species. Proceedings of the Helminthological Society of Washington, 50(2): 261-274.

Lichtenfels, J. R. and Pilitt, P. A. А (1991): redescription of Ostertagia bisonis (Nematoda : Trichostrongyloidea) and a key to species of Ostertagiinae with a lateral synlophe tapering from domestic ruminants in North America. Journal of the Helminthological Society of Washington, 58(2): 231-244.

Lichtenfels, J.R.; Pilitt, P.A. and Le Jambre, L. F. (1986): Cuticular ridge patterns of *Haemonchus contortus* and *Haemonchus placei* (Nematoda : *Trichostrongyloidea*). Proceedings of the Helminthological Society of Washington, 53(1): 94-101.

Lichtenfels, J.R.; Pilitt, P.A. and Hoberg, E.P. (1994): New morphological characters for identifying individual specimens of *Haemonchus* spp. (Nematoda : *Trichostrongyloidea*) and a key to species in ruminants of North America. Journal of Parasitology, 80(1): 107-119.

Luna, L.G. (1968): Manual of Histologic Staining Methods of the Armed Forces Institute of Pathology, 3rd edn. New York, McGraw-Hill Book Company.

Mckellar, Q.A.; Jackson, F.; Coop, R.L.; Jackson, E. and Scott, E. (1991): Effect of parasitism with Nematodirus battus on the of pharmacokinetics levamisole. netobimin. ivermectin and Veterinary Parasitology, 39(1-2): 123-36.

Mir, M.R.; Chishti, M.Z.; Rashid, M.; Dar, S.A.; Katoch, R.; Kuchay, J.A. and Dar, J.A. (2013): Point prevalence of Gastrointestinal Helminthiaisis in large Ruminants of Jammu, India. International Journal of Scientific and Research, 3(3): 2250-3153.

Muluneh, J.: Bogale, B. and Chanie. Μ. (2014): Major gastrointestinal Nematodes of small ruminants in Dembia District, Northwest Ethiopia. European Journal of Applied Sciences, 6(2): 30-36.

Nwosu, C.O.; Madu, P.P. and Richards, W.S. (2007): Prevalence and seasonal changes in population of gastrointestinal nematodes of small ruminants in zone the semi-arid of northeastern Nigeria. Veterinary Parasitology, 144(1-2): 118-24.

Owhoeli, O.; Elele, K. and Gboeloh, L.B. (2014): Prevalence of gastrointestinal helminthes in exotic and indigenous goats slaughtered in selected abattoirs in Port Harcourt, South-South, Nigeria. Chinese Journal of Biology, vol.2014: 1-8.

Pfukenyi, D.M.; Mukaratirwa, S.; Willingham, A.L. and Monrad, J. (2007): Epidemiological studies of parasitic gastrointestinal nematodes. cestodes and coccidian infections in cattle in the Highveld and lowveld communal Zimbabwe. grazing areas of Onderstepoort Journal of Veterinary Research, 74(2): 129-142.

Rauf Tak, I.; Ali Dar, Sh.; Shafi Dar, J.; Ganai, B.A.; Chishti, M.Z. and Ahmed, F. (2014): A brief study of morphology of *Haemonchus contortus* and its hematophagous behavior. Global Veterinaria, 13(6): 960-965.

Raza, M.A., Murtaza S., Bachaya H.A., Dastager G. and Hussain A. (2009): Point prevalence of Haemonchosis in sheep and goats slaughtered at Multan abattoir. The Journal of Animal & Plant Sciences, 19(3): 158-159.

Regassa, F.; Sori, T.; Dhuguma, R. and Kiros, Y. (2006): Epidemiology of gastrointestinal parasites of ruminants in Western Oromia, Ethiopia. International Journal of Applied Research in Veterinary Medicine, 4(1): 51-57.

Saminathan, M.; Gopalakrishnan, A.; Latchumikanthan, A.; Milton, A. A.P.; Aravind, M.; Dhama, K. and Singh, R. (2015): Histopathological and parasitological study of Blood-Sucking *Haemonchus contortus* infection in sheep. Advances in Animal and Veterinary Sciences, 3(2): 99.

Soulsby, E.J.L. (1982): Helminths, arthropods, and protozoa of domesticated animals. Seventh Ed. Lea & Febiger, Philadelphia, USA.

Sultan, K.; Desoukey, A.Y.; Elsiefy, M.A. and Elbahy, N.M. (2010): An abattoir study on the prevalence of some gastrointestinal helminths of sheep in Gharbia Governorate, Egypt. Global Veterinaria, 5(2): 84-87.

Sultan, K.; Elmonir W. and Hegazy Y; (2016): Gastrointestinal parasites of sheep in Kafrelsheikh governorate, Egypt: prevalence, control and public health implications. Beni-Suef University Journal of Basic and Applied Sciences, 5(1): 79-84.

Swapna, S. and Nithinya, R. (2015): Prevalence of gastrointestinal nematode and trematode parasites of cattle in Northern Kerala, India. International Journal of Science and Engineering, 3(2): 132-136.

Tehrani, A.; Javanbakht, J.; Jani, M.; Sasani, F.; solati, A.; Rajabian, M.; Khadivar, F.; Akbari, H.; and Mohammadian, M. (2012): Histopathological study of *Haemonchus contortus* in Herrik sheep abomasum, Iran. Journal Bacteriolgical Parasitology, 3(5): 144-149.

Vadlejch, J.; Lukešová, D.; Vašek, J.; Vejl, P.; Sedlák, P.; Čadková, Z.; Langrová, I.; Jankovská, I. and Salaba, O. (2014): Comparative morphological and molecular identification of *Haemonchus* species in sheep. Helminthologia, 51(2): 130 – 140.

Varadharajan, Α. and Vijavalakshmi, R. (2015): Prevalence and seasonal of gastrointestinal occurrence parasites in small ruminants of coastal areas of Tamil Nadu. International Journal of Scientific and Research Publications, 5(2): 159-160.

Walsh, A.; Younis, P.J. and Morton, (1995): J.M. The effect of ivermectin treatment of late pregnant dairy cows in south- west subsequent Victoria on milk production and reproductive performance. Australian Veterinary Journal, 72(9): 360.

Zajac, A.M. and Conboy, G.A. (2012): Veterinary clinical parasitology eighth Ed., John Wiley & Sons.

الملخص العربى

دراسات وصفية وبيولوجيةعلي الديدان الاسطوانية المعدمعوية فى المجترات

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اجريت هذه الدراسة للبحث عن إصابه المجترات بالديدان الإسطوانية المعدمعوية في ثلاثة مدن الاسماعيلية وبورسعيد ودمياط في الفترة من اكتوبر ٢٠١٦ الي سبتمبر ٢٠١٧ وتم فحص ١٣٠ عينة براز من الحيوانات الحية بصورة عشوائية وكذلك فحص ٢٠عينة من المعده الرابعة و٣٦ عينة من الامعاء الدقيقة من الحيوانات المذبوحة في المجازر ثم تم إجراء الصفة التشريحية لها للبحث عن الديدان الاسطوانية واظهرت النتائج إصابة الاغنام ب (ثلاثة أنواع من الديدان الإسطوانية في المعدة الرابعة و نوعين من الديدان الإسطوانية في الأمعاء الدقيقة) وكانت نسبة اصابة المجترات في عينات البراز ٢٦.١٥% وفي عينات المجزر (٣٠% في المعدة الرابعة و ١١.١١% في الأمعاء الدقيقة) وقد سجل وجود فروق معنوية في انتشار الطفيليات بين أعمار الحيوانات حيث أعلى نسبة إصابة في الحيوانات الصغيرة من عمر ٢-١٢ شهربنسبة إصابة تصل الى ٢٢. ٧١% في الأغنام و ٢٨. ١٤% في الابقار وكذلك نسبة انتشار الإصابة في ذكور الحيوانات اكثر من الإنات حيث في ذكور الأغنام وصلت النسبة الى ٦٠% وفي الإناث ٣٠% كما وجدت فروق معنوية في معدل الإصابة من حيث منشأ الحيوان حيث سجلت محافظة دمياط أعلى نسبة في معدل الإصابة فكانت النسبة ٤٧.٦% وتليها محافظة الاسماعيلية وسجلت ٢٩.١٦% ولم تسجل النتائج ظهور إصابة في محافظة بورسعيد وكذلك وجدت فروق معنوية في معدل الإصابة باختلاف شهور السنة حيث كان شهر سبتمبر اكثر الشهور انتشارا للمرض بنسبة ٢٦.٦٧% وكان اكثر الانواع انتشارا في المعدة الرابعة ديدان *الهيمونكس* بنسبة ٢٦.٦٧ ثم ديدان الأوسترتاجيا بنسبة ١٠% واخبرا ديدان التريكوسترونجيلوس بنسبة ٣.٣٣% اما في الامعاء فسجلت النتائج ديدان *الكوبيريا* بنسبة ١١.١١% وكذلك ديدان *النيماتوديرس* بنسبة ٥٦.٥% كما اشارت النتائج إلى وجود تغيرات نسيجية واضحة بوجود تنخر في الطبقة المخاطية وتثخن بالطبقة العضلية وفرط التتسج للخلايا الجداريه مع الخزب بالاضافه إلى وجود ارتشاح للخلايا الالتهابية متعددة الأشكال وتليف كامل لبعض الخلايا الغديه للمعدة الرابعة. وقد تم عمل اختبار تتاقص عدد البيض بإثنين من مضادات الديدان الإسطوانية وهما الأيفوماك سوبر و الألبندازول وتبين تغوق الأيفوماك سوبر على الألبندازول في علاج الديدان الإسطوانية في المجترات.