Egyptian Veterinary Medical Society of Parasitology Journal



Original Article

Molecular and Pathological Studies on *Sarcoptes scabiei* in Sheep in Ismailia Province, Egypt.

Mariam.A.Atteya¹, Mahi.A.Ghobashy², Wahba. A.A³, Eman M.Abouelhassan⁴

^{1,3}Department of Parasitology, Animal Health Research Institute;
² Department of Zoology, Faculty of Sciences, Suez Canal University, Ismailia, Egypt ⁴ Department of Parasitology, Faculty of Veterinary Medicine, Suez Canal University, Ismailia, Egypt;.

Corresponding author: Eman M.Abouelhassan, Email: <u>hassanemy@yahoo.com</u>

Tel.: 01280035453

Abstract:

Scabies is one of the important neglected tropical skin diseases which caused by the parasitic mite Sarcoptes scabiei. Scabies is usually detected in the developing countries. This study was designed to investigate the molecular characterization and pathological alterations induced by mite infestation in sheep at Ismailia province, Egypt. A total of 760 sheep aged from 3 months to 3 years were examined during the period from October 2017 to March 2019. Among them, 80 (10.5%) were infested with one species of mites Sarcoptes scabiei. Molecularly, the second internal transcribed spacer (ITS-2) of nuclear ribosomal DNA (rDNA) was as a genetic marker. TheITS-2 primer was amplified from individual mites by polymerase chain reaction (PCR) revealing the diagnostic specific band at 480 bp, nucleotides sequencing proved the species. Grossly, alopecia with rough leathery and corrugation of skin, erythema, crusts and pruritus were observed. The associated histological lesions were hyperkeratosis with crusting and thickening of the epidermis, acanthosis and vesiculation.

Key words: *Sarcoptes scabiei*, Alopecia, Crusts, Pruritus, Hyperkeratosis, Acanthosis, *ITS2*, PCR.

Introduction

Mange is a severe contagious disease and a major global health problem affecting humans and other mammals (Currier et al., 2011). Mites are the common cause of skin diseases in sheep. They can cause hypersensitivity disorders in animals. They may also cause life threatening anemia in young and/or weakened animals (Araujo et 1998), thev are microscopic al.. ectoparasites which can cause mild to chronic skin disease known as "Mange" in several hosts including domestic, farm and wild animals. It is an important emerging disease of wildlife and a well-recognized threat to the health and sometimes the existence of endangered or isolated wildlife populations throughout the world (Pence and Ueckermann, 2002).

Mange is characterized by a loss of hairs with itching and scabby eruptions in the affected animals, humans might be also affected (Currier *et al.*, 2011). Skin lesions usually begin with erythematous areas and develop to papules with crusts formation (Osman *et al.*, 2006).

Clinical signs observed in animals in acute infestations consisted of intense pruritus, erythematous eruptions, alopecia, and seborrhea (Burkhart al., 2000). et Sometimes, erythematous and alopecic areas as well as an intense pruritus appear with severe hair loss and hyperkeratosis (Rentería-Solís et al., 2014). Meanwhile, Pence et al. (1983) described that crusted plaques, alopecia, or absent hair and with/without crusts have been found in canines.

The infestation of scabies in four wild raccoon dogs, Histologically, epidermal hyperplasia showed, in the papillary dermis with acanthosis accompanying marked rete ridge formation, hyperkeratosis that was predominantly parakeratotic in focal areas, and subcorneally formed tunnels in which mites were evident. Some epidermal tunnels were covered with flattened parakeratotic cells (Eo et al., 2008). In the superficial dermis, infiltrates of cells in a perivascular pattern were detected. A marked acanthosis and hyperkeratosis, predominantly keratotic well as as multifocal mild inflammatory infiltrate, predominantly composed of neutrophils and occasional eosinophils, and superficial bacterial colonies were seen (Teodoro et al., 2018). Subcorneal pustules associated with a discrete lymphoplasmacytic infiltrate were observed in the skin and the superficial dermis of one of the canids (Teodoro et al., 2018).

Host-associated populations of S. scabiei taxonomically are divided into morphologically indistinguishable varieties with a high degree of host specificity and a low degree of cross infectivity. The varieties are named based on their host species: S. scabiei Var. hominis, S. scabiei Var. canis. Historically, genetic research on scabies has been extremely limited (Walton et al., 2004). This is primarily might be due to the difficulty in obtaining sufficient quantities of the mite and usable amounts of genetic material. The taxonomic status of mites of the genus Sarcoptes was clarified by Zahler et al. (1999) utilizing the second internal transcribed spacer (ITS-2) of the rRNA gene, and described the phenotypic characters, and investigated them in 23 isolates from different host species (pig, cattle, dog, fox, raccon and lynx). Phenotypic differences among isolates were observed. Concerning the genotypic difference between distinct groups, they observed that there was no correlation with the different host species or even with the geographic origin. These results supported the co-specificity of the mites investigated and proved the concept that the genus *Sarcoptes* consists of a single heterogeneous species.

Material and Methods

1. Study area and animals

A total of 760 sheep, aged from 3 months to 3 years screened for mites infestation during the period from October 2017 to March 2019 from different farms in Ismailia province (Kasassin, Elaabtal. Kantara, AbuAtwa, Sarabiom and Abuswear). The suspected animals (110/760) were examined following the standard procedures of skin scrapings, 80 suffered from Sarcoptic mange.

2. Examining and Sampling from the animals

Sheep were clinically examined to evaluate the health condition of the animals and to exclude any other disease affecting the health condition.

After clinical examination, suspected sheep were selected for parasitological investigation. Profound skin scrapings from the peripheral or the edges of lesions which were obtained from different areas of the body such as head, face, neck, ears, tail and trunk (scraping area varied from 1 to 2 .5 cm²) area of affected lesions, until the skin was bleeding slightly.

3. Parasitological preparation and identification

Scraped samples were treated with 5 ml of 10% of potassium hydroxide to dissolve tissue materials and heated for 5-10 min. Thereafter, samples were centrifuged at 1500 rpm for 4-5 min, and then the sediment was spread on a glass slide, microscopically examined under 10x magnification.

Permanent preparation of mite specimens

Mite specimens were mounted on glass slides from 70% alcohol after rinsing in

water using Berlese medium (Hoyer's) which is prepared according to (Baker & Wharton, 1959). Species identification of mites was determined morphologically according to Mellanby (1985).

4. Molecular identification

DNA extraction and PCR analysis:

A total of 17 specimens were utilized in this previously observed by study. light microscope, DNA was extracted from these specimens using the Genomic DNA Purification kit (Applied biotechnology). PCR amplification of the ITS-2 was done using primers RIB-18 and RIB-3 as described by Zahler et al. (1999). The PCR was done following the cycling condition: initial denaturation at 95°C for 5 minutes followed by 10 cycles of 92°C for 1min, 48°C for 1 minute and 72°C for 90 seconds. This step was followed by additional 32 cycles of 92°C for 1 minute, 54°C for 35 seconds and 72°C for 90 seconds, this was followed by a final extension at 72°C for 7 minutes. The amplification products from ITS-2 were separated on 1.6% agarose gel containing 0.4 µg/ml of ethidium bromide (Bio-Rad Laboratoies Inc., Hercules, CA) at 90 volts for 40-60 minutes, The PCR products were sent for sequencing. Sequences were amplified using primers the upstream primer RIB-18 5' -GGG CTG CAG TAT CCG ATG GCT TCG T-3'. and RIB-3 5 '- CGG GAT CCT TC (A,G) CTC GCC G(C,T)T ACT- 3'. **DNA Sequence analysis:**

PCR products were sequenced directly using Big Dye Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems) and an ABI 3130 Genetic Analyzer (Applied Biosystems). Sequences were assembled using the ChromasPro. The accuracy of data was confirmed by bi-directional sequencing. The obtained sequences were

EVMSPJ 2020; 16:50-62

aligned with each other and reference sequences of each gene using ClustalX to confirm the identification of *S. scabiei*. Maximum Parsimony analysis of taxa methods and in the MEGA X10.1 software was used to assess the phylogenetic relationship among different populations of *S. scabie* (Kumar *et al.*, 2018)

5. **Histopathological Preparations:** The skin of slaughtered sheep infested with Sarcoptes scabiei was collected and immediately fixed in 10% formalin, washed several times in 70% ethanol and then fixed in a mixture of 70% ethyl alcohol 95% and glycerin 5%. The specimens were then dehydrated in ascending grades of ethyl alcohol, cleared in xylene for two days, then washed in benzene for 10 minutes and embedded in three changes of pure paraffin wax. Serial transverse sections of skin, 5 microns-thick sections were cut and mounted on clean glass slides, stained in haematoxylin and eosin, cleared in xylene and mounted in Canada balsam (Bancroft and Bamble, 2008).

RESULTS

1.Clinical observation:

The initial lesions were localized accompanied by alopecia and were found on the body parts which had thin skin and less wool. Scabby lesions appeared as erythematous skin with irregular alopecic areas and scab formation Fig (1A&B).

2. Morphological identification:

The collected *Sarcoptes scabiei* were identified by its size, shape and morphology following up the identification key of Mellanby, (1985). Adult scabies mites are roughly round, ventrally flattened and dorsally convex tortoise-like bodies. Adult female scabies mites have four pairs of short legs (two pairs in front and two pairs behind), and its first and second pairs of legs well separated from the third and fourth pairs of legs Fig. (2A, B&C).

3. Molecular identification:

In this study, a total of 17 mite specimens were analyzed based on ITS2 PCR (Zahler *et al.*, 1999) (Fig 3), the specimens sequences had been blasted on the genebank and were identified as *Sarcoptes scabiei* with 99% identity.

Phylogenetic analysis:

The phylogenetic analysis was performed using MEGA X10.1 software and the tree was constructed using maximum parsimony analysis of taxa methods (Fig: 4). The sequenced samples clustered with *Sarcoptes scabiei* species from Iran and Egypt. There are low degree of sequences variation observed with them since they all share the same ancestor.

4. Pathological lesions produced by the mite:

Grossly, alopecia with rough leathery and corrugation of skin, erythema, crusts and pruritus were observed. Poor body condition, sunked eyes, and extensive areas of alopecia were seen at the skin of slaughtered sheep obtained from the abattoir, and red smooth areas appeared with thick, irregular, opaque skin and yellow-brownish crusts that detached easily. These areas were observed on the head, trunk and abdomen.

Histopathological description:

Microscopically, it was characterized by acanthosis, hyperkeratosis, the formation of hyperplastic rete-pegs (Fig 5, A; 5, C), destruction of dermis and epidermis (Fig 5, B), hyperplastic changes in sebaceous glands, sweet gland and hair follicular cells, pyogranuloma in papillary layer and hair follicles and infiltration of neutrophils, eosinophils, lymphocytes and few macrophages (Fig 5, D). **Figure legends**



Figure (1): (A) Adult sheep infected with *Sarcoptic scabiei* mites suffer from damaged wool in abdomen area; (B): Adult male sheep infested with *Sarcoptic scabiei* mites on the ear and abdomen area.

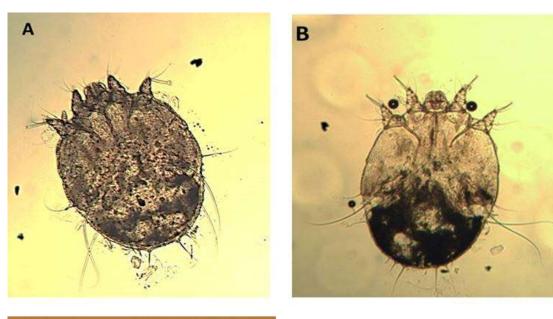




Figure (2): (A, B): Light microscopy (LM) of fresh specimens Sarcoptes scabiei adult (A): dorsal view (B): ventral view (C): (LM) of permanent preparation Sarcoptes scabiei adult

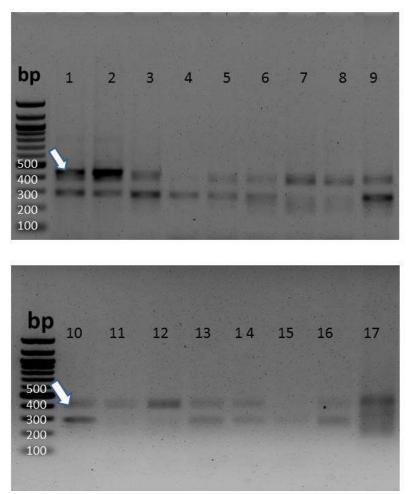
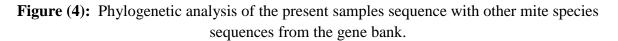


Figure (3) Analysis of ITS-2 PCR products of *Sarcoptes scabiei* by agarose gel electrophoresis. Left Lane represents 100 bp (base pair) DNA ladder plus marker, **lanes from** (1to17) represent PCR product for DNA extraction samples of *Sarcoptic scapiei* isolated from sheep with product size 480 bp.





Maximum Parsimony analysis of taxa

The evolutionary history was inferred using the Maximum Parsimony method. The most parsimonious tree with length = 1060 is shown. The consistency index is 0.807183 (0.764977), the retention index is 0.588710 (0.588710), and the composite index is 0.475197 (0.450349) for all sites and parsimony-informative sites (in parentheses). The MP tree was obtained using the Subtree-Pruning-Regrafting (SPR) algorithm (pg. 126 in ref. [1]) with search level 0 in which the initial trees were obtained by the random addition of sequences (10 replicates). This analysis involved 9 nucleotide sequences. There were a total of 467 positions in the final dataset. Evolutionary analyses were conducted in MEGA X.

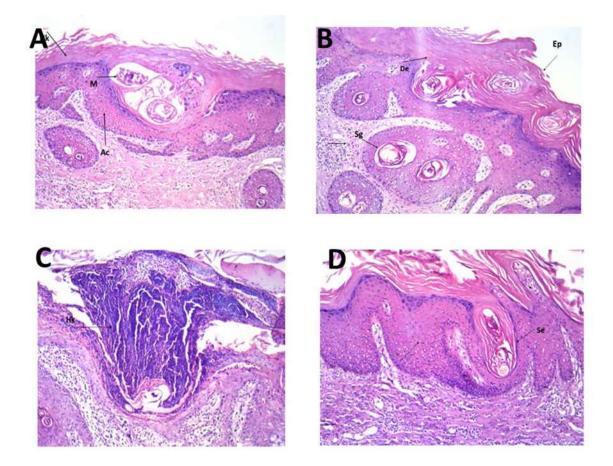


Figure (5) (A): Sarcoptic mange in sheep. Histological aspect of the skin with *Sarcoptes scabiei*, in which are evidenced idiosoma (body of mite) (M) associated with predominantly orthokeratotic hyperkeratosis (Hk) and acanthosis (Ac). X100; (B): Destruction of dermis (De) and epidermis (Ep), hyperplastic changes in sebaceous glands (Sg) and infiltration of neutrophils, eosinophils, lymphocytes and few macrophages (arrow). X10; (C): Epidermal covering lined by hyperplastic stratified squamous epithelium showing mild papillomatosis, moderate acanthosis, marked hyperkeratosis (Hk) with focal hypergranulosis and parakeratosis. X100; (D): Microscopic figure characterized by acanthosis (arrow), showing changes in sweet gland (Se). X100

Discussion

The present study showed that infestations *scabiei* in sheep of Sarcoptes was potentially significant. The result demonstrated that the induced lesions of the mite in host tissues produced irritations which led to itching and scratching. The resulting inflammation of the skin is accompanied bv exudate which an becamecoagulative and formed crusts on the surface and is further characterized by an excessive keratinization and proliferation of connective tissues.Progressively, the skin became much thickened and wrinkled.

In some animals, deaths occurred possibly due to the malnutrition, as the affected animals spent less time feeding because of the intense pruritus, and the severity of the lesions which can lead to ulcers formation, myiasis, secondary bacterial infections, toxemia (Mauldin & Peters-Kennedy, 2016).

Currently, the molecular identification based on DNA sequences was applied to overcome the fact that the morphological identification is insufficient for the accurate detection of the species (Abouelhassan et al., 2019). The sequencing analysis based on the amplification of the ITS-2 rDNA as well as the phylogenetic analysis proveded that the examined mite species were Sarcoptes scabiei. This result was in concordance with those of Zahler et al. (1999) and Gu and Yang (2008). The former reported a very little genetic variation among sarcoptic mites collected from different hosts and geographic locations, while Gu and Yang (2008) could not differentiate sarcoptes mites among different hosts in China. Although Berrilli et al. (2002) detected some genetic variability between individual mites. The sequence variations were randomly distributed in different hosts from several locations, thus, resulting in an indistinct geographic or host-specific clustering.

in Egypt Sarcoptes scabiei was recorded by Yassin (2011) in Egyptian buffaloes at Giza governorate, in addition to Mazyad et al., (2001) who reported the presence of sarcoptes mites in man and sheep in North Sinai, The present study revealed that there is no genetic variation in the sarcoptic mites collected from different sheep farms in the same geographic area and there is little genetic variation comparing with the other sequences from the genebank of the collected mite species from different geographic locations Egypt. in Furthermore, the present findings went parallel with those Amer et al. (2014) who used ITS2 sequence analysis for the sarcoptes mites derived from different hosts (water buffalo, cattle, sheep, and rabbits) from farms at K afr El Sheikh, Egypt. He reported very little genetic variation in sarcoptic mites from different hosts species with ITS2 and geographic locations sequence analysis.

Herein, histopathological alterations appeared consisted of hyperkeratosis with crusting and thickening of the epidermis, acanthosis, vesiculation and mites in the stratum corneum. Dermal affections were intradermal proliferation of connective tissue, edema in the papillary layer and severe degenerative and necrotic changes of hair the follicles. Those findings corresponded to the classic description of sarcoptic mange in dogs (Morris, 1996; Teodoro et al., 2018). In addition, the resulted characteristic pruritic ectoparasitism was the appearance of hyperplasia and infiltration of eosinophils (Morris, 1996; Teodoro *et al.*, 2018). The occurrence of numerous mast cells in the affected dermis was consistent with the pathogenesis of sarcoptic mange, which is largely associated with hypersensitivity to the mites (Skerratt, 2003). Those features agreed with the effect of sarcoptic mange in domestic and wild mammals (Nimmervoll *et al.*, 2013). Interestingly, the cellular response was also clear (Eo *et al.*, 2008, Nimmervoll *et al.*, 2013 and Teodoro *et al.*, 2018).

Conclusion:

Mange is a contagious and debilitating skin disease of sheep and a notable problem in Ismailia province. It could be concluded that the molecular identification of the mite species, based on DNA sequences, is an alternative and demanded tool for the accurate identification of mite species to overcome the difficulties associated with the morphological identification, Further studies are required to choose good genetic marker to help in identification of mite species.

References

Abouelhassan E.M.,ElGawady H.M, Abdelaal A.A., El-GayarA. K. and

Esteve-Gassent M.D. (2019). Comparison of Molecular tick identification based on 16S rDNA, 12S rDNA, ITS1 and ITS2. Journal of Arthropds borne diseases 13(2):153-164.

Amer S., Said E., Taher A.M., Abd El Naby Y., Jianbin R., Dawn F., Yaoyu X.L. (2014). "Preliminary molecular characterizations of Sarcoptes scaibiei (Acari: Sarcoptidae) from farm animals in Egypt." PloS one 9(4): e94705.

Araujo, F.R., Silva, M.P., Lopes, A.A., Ribeiro, O.C., Pires, P.P., Carvalho, C.M., Balbuena, C.B., Villas, A.A. and Ramos, J.K. (1998). Comunidades vegetais do Parque Nacional da Restinga de Jurubatiba, pp. 39-62. In: F. A. Esteves (ed.), Ecologia das lagoas costeiras do Parque Nacional de Jurubatiba e do Município de Macaé, RJ. NUPEM/UFRJ, Macaé, RJ.

Baker, E. W. and Wharton, G. W. (1959). An introduction to acarology 3rd Edit., the Macimillan Co. N.York.

Bancroft, J.D. and Gamble, M. (2008) Theory and Practice of Histological Techniques. 6th Edition, Churchill Livingstone, Elsevier, China.

Berrilli E S., D'Amelio E.L., Rossi. (2002). Ribosomal and mitochondrial DNA sequence variation in Sarcoptes mites from different hosts and geographical regions Parasitol Res 88: 772–777.

Burkhart C.G., Burkhart C.N. & Burkhart K.M. (2000). An epidemiologic and therapeutic reassessment of scabies. Cutis 65(4):233-240.

Currier R.W., Shelley F.W., and Bart J.C. (2011). "Scabies in animals and humans: history, evolutionary perspectives, and modern clinical management." Annals of the New York Academy of Sciences 1230(1).

Eo K., Kwon O., Shin N., Shin T., and Kwak D. (2008). Sarcpoptic mange in wild raccon dogs (*Nyctereutes Procyonoides*) in korea. Zoo and Wildlife Medicine 39(4): 671–673.

Gu X. and Yang G. (2008). A study on the genetic relationship of mites in the genus *Sarcoptes* (Acari: Sarcoptidae) in China. International Journal of Acarology, Volume 3 4.

-Kumar S., Stecher G., Li M., Knyaz C., and Tamura K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. Molecular Biology and Evolution 35:1547-1549.

Mazyad S., Sanad E., Morsy T. (2001). Two types of scab mites infesting man and sheep in North Sinai. J Egypt Soc Parasitol 31: 213–222.

Mauldin E.A. and Peters-Kennedy J. (2016). Kennedy and Palmer's Pathology of domestic animals. 6th ed. Elsevier, Edinburgh.

Mellanby K. 1985. Biology of the parasite. In: Orkin M, Maibach HI (eds). Cutaneous infestations and insect bites. New York. Marcel Dekker. pp 9-18.

Morris D.O. (1996). A histomorphological study of sarcoptic acariasis in the dog: 19 cases. Journal of the American, Animal Hospital Association; 32: 119–24.

Nei M. and Kumar S. (2000). Molecular Evolution and Phylogenetics. Oxford University Press, New York.

Nimmervoll H., Hoby S., Robert N., Lommano E., Welle M. and Ryser-Degiorgis M.P. (2013). "Pathology of Sarcoptic mange in red foxes (*Vulpes Vulpes*): Macroscopic and histologic characterization of three disease stages". Journal of Wildlife Diseases, 49: 91–102.

Osman S.A., Hanafy A. and Amer S.E. (2006): Clinical and therapeutic studies on mange in horses. Vet Parasitol., 10; 141(1-2):191-195.

Pence D. and E. Ueckermann (2002). Sarcoptic mange in wildlife. Revue Scientifique Technique-Office International des Epizooties

Pence D.B., Windberg L.A., Pence B.C.
& Sprowls R. (1983). The epizootiology and pathology of sarcoptic mange in coyotes, *Canis latrans*, from South Texas.
J. Parasitol. 69(6):1100-1115.

Rasero R., Ryser Degiorgis M.P., Nimmervoll H., Zhu X.Q. and Rossi L. (2009). Is ITS-2 rDNA suitable marker for genetic characterization of Sarcoptes mites from different wild animals in different geographic areas? Veterinary Parasitology 159 181–185.

Rentería-Solís Z., Min A.M., Alasaad S., Müller K., Michler F.U., Schmäschke R., Wittstatt U., Rossi L. & Wibbelt G. (2014). Genetic epidemiology and pathology of raccoon-derived Sarcoptes mites from urban areas of Germany. Med. Vet. Entomol. 28(Suppl.1):98-103.

Skerratt L.F. (2003). Cellular response in the dermis of common wombats (*Vombatus ursinus*) infected with *Sarcoptes scabiei* var. *wombati*. J. Wildl. Dis. 39: 193–202.

Soulsby E. (1986). Helminths, arthropods and protozoa of domestic animals, 7 edn. Bailliere M. Tindall, London.

Teodoro G.W., Pâmela A. L., Patrícia C. S., Ivam M.J., Mary S. V., Flademir W. and Angelica T.B. (2018). Sarcoptic mange (*Sarcoptes scabiei*) in wild canids (*Cerdocyon thous*) Pesq. Vet. Bras. 38(7):1444-1448.

Walton S.F., Holt D.C., Currie B.J. and Kemp D.J. (2004). Scabies: New future for a neglected disease. Advances in Parasitology 57: 309-376.

Yassin M. (2011). Mange mites causing scabies in Egyptian buffaloes at Giza Governorate. Egypt J Egypt Soc Parasitol 41: 55–64.

Zahler M., Essig A., Gothe R. and Rinder H. (1999). "Molecular analyses suggest monospecificity of the genus Sarcoptes (Acari: Sarcoptidae) 1." International journal for parasitology 29(5): 759-7

الجرب من اهم الامراض الطفيلية التي تصيب الاغنام في مصر و هو واحد من امراض الجلد الاستوائية المهملة فى البلدان النامية، صممت هذه الدراسة لتقصي الخصائص الجزيئية والتغيرات المرضية التي يسببها الحلم في الاغنام بحافظة الاسماعيلية بمصر و قد تم فحص ٢٠ خروفا تتراوح اعمارهم من ٣ اشهر الى ٣ سنوات خلال الفترة من اكتوبر ٢٠١٧ الي مارس ٢٠١٩ وتبين اصابة عدد ٨٠ خروفا بنسبة ٥٠٠ % بنوع واحد من حلم الجرب

وقد تم التعرف علي النوع ميكروسكوبيا و عند فحص الحمض النووي للعينات عن طريق تفاعل انزيم البلمرة المتسلسل با ستخدام الجين الداخلي المنسوخ

وقد لو حظ تغيرات في الجلد المصاب بالطفيل مثل ظهور تصلب وقشور واختفاء للشعرمع التغيرات المرضية التى وجدت باستخدام الميكروسكوب .